Nogo-A Downregulation Improves Insulin Secretion in Mice

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Type 2 diabetes (T2D) is characterized by β-cell dysfunction and the subsequent depletion of insulin production, usually in a context of increased peripheral insulin resistance. T2D patients are routinely treated with oral antidiabetic agents such as sulfonylureas or dipeptidyl peptidase-4 antagonists, which promote glucose- and incretin-dependent insulin secretion, respectively. Interestingly, insulin secretion may also be induced by neural stimulation. Here, we report the expression of Nogo-A in β-cells. Nogo-A is a membrane proteoglycan that inhibits neurite outgrowth and cell migration in the central nervous system. We observed that Nogo-A-deficient mice display improved insulin secretion and glucose clearance. This was associated with a stronger parasympathetic input and higher sensitivity of β-cells to the cholinergic analog carbachol. Insulin secretion was also improved in diabetic db/db mice treated with neutralizing antibody against Nogo-A. Together, these findings suggest that promoting the vagal stimulation of insulin secretion through the selective inhibition of Nogo-A could be a novel therapeutic approach in T2D. Diabetes 62:1443–1452, 2013

The relative or absolute lack of insulin is responsible for diabetes. In type 1 diabetes, β-cell loss is due in most cases to an autoimmune reaction, but not exclusively (1). In type 2 diabetes (T2D), increased peripheral insulin resistance challenges the functional β-cell mass; after an initial attempt at overriding the increased insulin demand, the number of cells that produce insulin declines progressively. Glucose entry into cells is regulated by insulin, whose secretion from β-cells is tightly coordinated by different secretagogues. Insulin secretion is initiated by the cholinergic parasympathetic stimulation of β-cells (the so-called “cerebral phase”) and subsequently potentiated during the enteric “absorptive phase” (2). In response to mechanical and chemical stimulation along the digestive tract, the intestinal incretin hormones glucagon-like peptide-1 (GLP-1) and gastric inhibitory peptide (GIP) potentiate insulin secretion (GIP) potentiate insulin secretion directly and indirectly, through neuronal stimulation (the “incretin effect”) (3–5). Progressively, nutrient absorption and increased blood glucose stimulate insulin secretion directly (post-absorptive phase) (6). Altogether, different secretagogues act synergistically and trigger the adequate biphasic release of insulin from β-cells, primed by cholinergic stimulation (7). These secretagogues reach islet endocrine cells through the vascular and neural networks. Pancreas innervation consists of parasympathetic (vagus nerve) and sympathetic efferent fibers and afferent sensory fibers (splanchnic nerve), and of intrapancreatic parasympathetic ganglion cells. The vagal input stimulates the secretion of insulin and other islet hormones, such as pancreatic polypeptide (PP) via cholinergic (i.e., mediated by acetylcholine) and noncholinergic mechanisms (8–10). Sympathetic postganglionic terminal nerves release noradrenaline or other peptides on endocrine cells; this represses insulin and somatostatin secretion while promoting glucagon release (11). The afferent sensory fibers innervate the periphery of islets and release calcitonin gene-related peptide (CGRP), among other peptides (12,13).

β-Cells and neurons share numerous features. They are electrically excitable, release mediators in response to membrane depolarization, and extend neurite-like processes (14). In addition, β-cells express many neuronal proteins (14,15), such as the neurotransmitter γ-aminobutiric acid (GABA) (16,17) or the synaptic cell-surface molecules neurexin, neuroligin, and SynCAM (18,19). Among them, neurexin and neuroligin have been shown to participate in insulin secretion (18,19).

Nogo-A is a high-molecular-weight membrane protein mostly expressed in the central nervous system (CNS), oligodendrocytes, and subsets of neurons (20,21), as well as other tissues, such as skeletal muscle (22). Nogo-A restricts neuronal regeneration in injured adult spinal cord and brain and limits plastic rearrangements and functional recovery after large CNS lesions, such as after spinal cord dorsal hemisection (23–25). In the intact CNS, Nogo-A appears to have a stabilizing and controlling role in axonal sprouting and cell migration (26–28). Cytoskeletal regulators, such as Rho GTPases or coflin, mediate the axonal and neurite growth inhibitory action of Nogo-A (28,29). Nogo-A and its receptor (NgR) are also found in synapses, where they may influence their stability and function (30–32).

Here we show that Nogo-A is expressed in pancreatic islets. We thus explored its potential role on endocrine pancreas function using mice lacking the two active Nogo-A alleles (33,34), which were challenged with different insulin secretagogues. Compared with wild-type animals, Nogo-A knockout (KO) mice presented increased insulin secretion, resulting in higher glucose clearance. This enhanced insulin release resulted from a higher pancreatic parasympathetic input on islets and from a higher sensitivity of β-cells to cholinergic and GLP-1 stimulation. We

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obtained similar results, i.e., improved insulin secretion associated with a higher responsiveness of β-cells, in diabetic db/db mice treated for a short period with neutralizing antibody against Nogo-A. Together, these observations reveal that Nogo-A is implicated in pancreatic endocrine function and thence in the control of glucose homeostasis.

RESEARCH DESIGN AND METHODS

Mice. Nogo-A−/− KO mice were described previously by one of us (33). In all experiments, C57BL/6J Nogo-A KO male mice were compared with age-matched C57BL/6J wild-type males. Genotyping was performed by PCR from genomic DNA isolated from tail biopsies using M58 (TGCTTGGATACTTCAAGTAGTCC) and M101 (AGTTGATCACCAGTCGAC) primers for wild-type Nogo-A allele (1.4-kb band), and M58 and M63 (CCCTACCCGGTTAGAATATCGAAGG) primers for the Nogo-A deleted allele (1.2-kb band). Five-week-old male db/db homozygous mice (C57BLKs background; BKS, Cg-Dock7<sup>-/-</sup> LeprdB strain) were purchased from Charles River Laboratories (L’Arbresle, France), for the treatment with neutralizing anti-Nogo-A 11C7 mouse antibody (25). Pdx1-GFP (35) and RIP-DTR (36) mice were previously described. Animals were maintained in a temperature-controlled room, on a 12-h light-dark cycle, and fed standard rodent chow ad libitum. The Directional Secretion Center of the Canton de Genève approved the study.

Islet isolation. Mice were killed and their pancreas exposed. After clamping at the porta hepatitis (transverse fissure of the liver), the main pancreatic duct was cannulated with a 27-gauge butterfly needle and retrogradually injected with 2 mL of collagenase XI (2 mg/mL in Hanks’ balanced salt solution) (Sigma-Aldrich). After dissection, the pancreas was digested in 3 mL of collagenase XI solution for 15 min at 37°C and finally disrupted upon vigorous shaking. Islets were then purified on a Histopaque 1119 (Sigma-Aldrich) gradient, washed three times with Hanks’ 1× BSA 0.1%, and finally “fished” (i.e., handpicked) twice to increase purity.

Fluorescence-activated cell sorter analysis. For fluorescence-activated cell sorter (FACS) sorting of GFP<sup>+</sup> cells, isolated Pdx1-GFP islets were dissociated into a single-cell suspension with trypsin 0.25%. Live β-cells were isolated as DAPI-GFP<sup>+</sup> cells by FACS with MoFlo (Dako-Cytomation). Islets were weighted and then rinsed in cold PBS and finally disrupted upon vigorous shaking. Islets were then purified on a Histopaque 1119 (Sigma-Aldrich) gradient, washed three times with Hanks’ 1× BSA 0.1%, and finally “fished” (i.e., handpicked) twice to increase purity.

Fluorescence-activated cell sorter analysis. For fluorescence-activated cell sorter (FACS) sorting of GFP<sup>+</sup> cells, isolated Pdx1-GFP islets were dissociated into a single-cell suspension with trypsin 0.25%. Live β-cells were isolated as DAPI-GFP<sup>+</sup> cells by FACS with MoFlo (Dako-Cytomation). Islets were weighted and then rinsed in cold PBS and finally disrupted upon vigorous shaking. Islets were then purified on a Histopaque 1119 (Sigma-Aldrich) gradient, washed three times with Hanks’ 1× BSA 0.1%, and finally “fished” (i.e., handpicked) twice to increase purity.

Immunofluorescence and immunohistochemistry. Collected pancreata were weighed and then rinsed in cold PBS and fixed overnight at 4°C in paraformaldehyde 4%. Tissues were dehydrated, embedded in paraffin, and sectioned at 5 μm with a microtome. The primary antibodies used for immunofluorescence were mouse antiglucagon (1/1,000, Sigma-Aldrich), guinea pig anti-insulin (1/400; Dako), rabbit anti-PP (1/200; Bachem), rabbit anti-somatostatin (1/200; Dako), rabbit anti-Brdu (1/500; gift from Christopher Wright, Vandebilt University, Nashville, TN), rabbit anti-β-Tubulin (1/1,000, Sigma-Aldrich), rabbit anti-NeuN (1/1,000, BCLC), and rabbit anti-Glut2 (1/200; gift from Bernard Thorens, Center for Integrative Genomics, University of Lausanne, Lausanne, Switzerland). For immunofluorescence experiments, the pancreata were sectioned at 5 μm with a microtome. Tissues were fixed with cold PBS and acetic acid (1:3) and then permeabilized with cold PBS containing 0.05% Triton X-100. Tissues were then incubated at 4°C overnight with the following primary antibodies: rabbit anti-glucagon (1/1,000, Sigma-Aldrich), rabbit anti-PP (1/100; Bachem), rabbit anti-somatostatin (1/200; Dako), rabbit anti-insulin (1/500; gift from Christian Wright, Vandebilt University, Nashville, TN), rabbit anti-BrdU (1/500; Dako), rabbit anti-β-Tubulin (1/1,000, Sigma-Aldrich), and rabbit anti-Glut2 (1/200; gift from Bernard Thorens, Center for Integrative Genomics, University of Lausanne, Lausanne, Switzerland).

In vivo administration of neutralizing anti-Nogo-A 11C7 antibody. Neutralizing anti-Nogo-A 11C7 mouse antibody (Novaris) and nonspecific anti-BrdU mouse antibody (AbSerotec) were intravenously infused once per day for 5 days at a total dose of 2 mg/kg. Tissues were then fixed with cold PBS and acetic acid (1:3) and then permeabilized with cold PBS containing 0.05% Triton X-100. Tissues were then incubated at 4°C overnight with the following primary antibodies: rabbit anti-glucagon (1/1,000, Sigma-Aldrich), rabbit anti-PP (1/100; Bachem), rabbit anti-somatostatin (1/200; Dako), rabbit anti-insulin (1/500; gift from Christian Wright, Vandebilt University, Nashville, TN), rabbit anti-BrdU (1/500; Dako), rabbit anti-β-Tubulin (1/1,000, Sigma-Aldrich), and rabbit anti-Glut2 (1/200; gift from Bernard Thorens, Center for Integrative Genomics, University of Lausanne, Lausanne, Switzerland).

RESULTS

Nogo-A is expressed in endocrine cells of the pancreas. In the adult mouse pancreas (2-month-old males), we detected by RT-PCR the expression of Rtn4a (Nogo-A transcript) in extracts from isolated islets of Langerhans (Fig. 1A). Rtn4a transcripts were also found in extracts from isolated β-cells but not from γ-cells isolated (data not shown). Islet cells isolated from Pdx1-GFP transgenic mice (35) (Fig. 1A). Rtn4a expression in β-cells was confirmed by deep sequence analysis performed on RNA extracts of β-cell–depleted islets obtained from RIP-DTR transgenic mice treated with diphertheria toxin (36) (Supplementary Fig. 1A). Finally, islet expression of Nogo-A at the protein level was observed using peroxidase-conjugated anti-rabbit IgGs (1/5,000; Promega). Bands were visualized by enhanced chemiluminescence (ECL; Amersham) according to the manufacturer’s instructions.

Statistical analysis. All results were reported as mean ± SEM. Groups were compared with nonparametric tests (one tailed Mann-Whitney), reported as P values. All tests were performed using the GraphPad Prism software.
by Western blot (Fig. 1B). We observed Rtn4a transcripts in human and rat islets as well (Supplementary Fig. 1B).

**Nogo-A KO mice display increased insulin secretion and lower glycemia, yet they have a normal β-cell mass.** The pancreatic expression of Nogo-A led us to wonder whether Nogo-A regulates insulin secretion, like neurelin (18). Body weight as well as the pancreas-to-body weight ratio was unchanged in Nogo-A KO mice (body weight: wild type 28.47 ± 0.86 g, KO 23.82 ± 1.89 g; n = 10, P = NS; pancreas-to-body weight ratio: wild-type 0.99 ± 0.05%, KO 0.86 ± 0.09%; n = 5, P = NS) (Fig. 2A and B). We measured plasma values of glucose, insulin, and glucagon in fasted and random-fed conditions. After 16-h fasting, plasma glucose, insulin, and glucagon levels were normal in KO mice (Fig. 2C–E). However, in a random-fed situation (a period during which insulin secretion is stimulated by glucose, gluconeeritins, and neurotransmitters), KO animals had significantly less blood glucose (wild type 10.66 ± 0.35 mmol/L, KO 8.48 ± 0.31 mmol/L; n = 5, P = 0.04), associated with increased insulinemia (wild type 0.50 ± 0.02 μg/L, KO 1.23 ± 0.28 μg/L; n = 6, P = 0.04) (Fig. 2C and D), but normal glucagonemia (wild type 60.79 ± 4.62 pg/L, KO 66.54 ± 13.29 pg/L; n = 6, P = NS) (Fig. 2C–E).

At the histological level, islet number, structure, size, and shape and β-cell mass were normal in Nogo-A KO mice (Supplementary Fig. 2A–E and not shown). The total pancreatic glucagon content was unchanged (wild type 338.07 ± 8.92 pg, KO 344.02 ± 4.96 pg; n = 9–11, P = NS) (Supplementary Fig. 2G), and the pancreatic insulin content was slightly increased (wild type 554.65 ± 54.54 ng, KO 763.51 ± 37.67 ng; n = 5, P < 0.05), thus suggesting that insulin content per β-cell was somewhat augmented (Supplementary Fig. 2F).

These observations reveal that although Nogo-A KO mice have a normal β-cell mass, they exhibit lower glycemia associated with higher insulin secretion in the random-fed condition, but without overt hypoglycemia. **Cholinergic-dependent glucose-stimulated insulin secretion is enhanced in Nogo-A KO mice.** In order to study the enhanced insulin secretion, we performed GTTs in response to different secretagogues (glucose alone or supplemented with carbachol) and after different routes of administration (intraperitoneal or intravenous injection). Besides increased plasma glucose and cholinergic stimulation (carbachol), insulin secretion from β-cells is also induced in response to the stimulated parasympathetic nervous system upon intraperitoneal injection of glucose (41). Overnight-fasted Nogo-A KO animals showed improved glucose clearance after intraperitoneal glucose administration (2 g/kg), as revealed by a decreased area under the curve (wild type 709.87 ± 72.73 mmol/L/min, KO 378.95 ± 31.17 mmol/L/min; n = 4, P = 0.01) (Fig. 3A). The stimulated insulin secretion was significantly higher 1 h after glucose injection in Nogo-A KO animals (Fig. 3B). Since insulin sensitivity was comparable in KO and wild-type mice (Fig. 3C), the faster glucose clearance in the former was thus due to a higher insulin secretion. We then explored the means by which insulin secretion is potentiated in Nogo-A KO mice. Therefore, after overnight fasting, animals were exposed to an intravenous bolus of either glucose alone (1 g/kg) (Fig. 3D and E) or glucose supplemented with the cholinergic analog carbachol (0.53 μmol/L), so as to mimic the vagal stimulation (Fig. 3F and G). Although control and KO mice similarly corrected the induced hyperglycemia by secreting comparable amounts of insulin in response to glucose (Fig. 3D–E), insulin secretion was higher in the KO group when animals were treated additionally with carbachol (Fig. 3F and G). Taken together, these observations suggest that in the absence of Nogo-A activity, the cholinergic vagal stimulation on β-cells triggers a stronger release of insulin.

**The increased insulin secretion in Nogo-A KO mice is associated with higher parasympathetic input on islet cells.** We reasoned that the enhanced insulin secretion in Nogo-A KO mice might rely upon higher islet parasympathetic input or increased β-cell responsiveness, or both. The density of parasympathetic (VACH'T⁺), sympathetic (VMAT²⁺), and sensory (CGRP⁺) fibers was normal in Nogo-A KO pancreas (not shown). We assessed the vagal stimulation on β-cells through the administration of 2DG (50 mg/kg; intravenously). 2DG is an unmetabolizable glucose analog that blocks intracellular glucose utilization, thus inducing neuroglycopenia and subsequently stimulating the vagus nerve, which promotes islet hormone secretion (12,42). We estimated the parasympathetic input on islets by measuring the fasting plasma PP, whose secretion is strictly controlled by parasympathetic stimulus (43,44). Whereas plasma PP was identical in the two groups of mice before 2DG injection, 30 min after 2DG, this value was almost doubled in Nogo-A KO mice (wild type 101.08 ± 11.45 pg/mL, KO 187.09 ± 15.33 pg/mL; n = 4, P < 0.05), suggesting an increased parasympathetic input (Fig. 4A). In parallel, we measured the postprandial plasma glucocerebrosidase, before and after 2DG injection. GLP-1 was undetectable, as previously reported (45), yet we could determine that plasma GIP values were lower in Nogo-A KO mice, irrespective of 2DG treatment, which correlates with their lower glycemia (Fig. 2C) (before 2DG: wild type 58.05 ± 4.90 pg/mL, KO 34.86 ± 3.08 pg/mL, P < 0.005; 15 min after 2DG: wild type 51.90 ± 4.65 pg/mL, KO 36.55 ± 4.19 pg/mL, P < 0.05, n = 5) (Fig. 4B). Combined, the above observations corroborate that the vagal stimulation of insulin secretion is reinforced by a higher parasympathetic tone in Nogo-A KO mice.

Using isolated islets, we assessed the responsiveness of β-cells to various secretagogues downstream of the parasympathetic stimulation. Groups of 10 islets with similar insulin content from control and Nogo-A KO mice responded similarly to various concentrations of glucose...
Accordingly, the expression of β-cell machinery genes (Insulin1, Insulin2, Slc2a2, GK, MafA, Nkx6.1, Ptx1, NeuroD1, Kcnj11, PC1, Gpr35, Htr7, Slc24a, and Trpm2) was fundamentally not altered, even though one gene, Trpm2, a regulator of insulin secretion (46), was upregulated in Nogo-A KO animals (Supplementary Fig. 3). Islets were then submitted to different glucose concentrations, supplemented with carbachol (100 μmol/L). In this situation, insulin secretion was increased in Nogo-A KO islets, as compared with control islets (1.4 mmol/L + CC: wild type 0.070 ± 0.008%, KO 0.195 ± 0.025%, P < 0.005; 8.4 mmol/L + CC: wild type 0.170 ± 0.025%, KO 0.287 ± 0.036%, n = 7–13, P < 0.05) (Fig. 4D). Finally, islets were incubated with glucose (8.4 mmol/L) supplemented with GLP-1 (100 nmol/L). Insulin secretion in response to glucose + GLP-1 was similar between wild-type and KO islets (Fig. 4E). These observations reveal that isolated Nogo-A KO islets secrete more insulin specifically in response to glucose supplemented with carbachol.

Diabetic db/db mice display improved insulin secretion after administration of neutralizing anti-Nogo-A antibody. The constitutive inactivation of Nogo-A confers to metabolically healthy mice the ability to more efficiently correct the induced hyperglycemia thanks to an improved insulin secretion. In order to explore the relevance of these observations to diabetes, we investigated the capacity of neutralizing anti-Nogo-A monoclonal antibody (termed 11C7) to promote insulin secretion in recent-onset T2D animals. Five-week-old diabetic db/db mice (i.e., lacking the leptin receptor) were treated with either anti-Nogo-A or control (anti-BrdU) antibody, as previously described (23). 11C7 was previously reported to inhibit Nogo-A in the CNS of adult rats, where it enhances the sprouting and regrowth of injured spinal cord axons (23). In brief, mice received two intravenous injections of 4.9 mg antibody (245 mg/kg), either 11C7 or control antibody, during a 2-week period. One week after the second injection, insulinemia in random-fed db/db mice treated
FIG. 3. Improved insulin secretion in Nogo-A KO mice in response to simultaneous glucose and nervous stimulation. A: Blood glucose (mmol/L) after intraperitoneal injection of 2 g/kg glucose is cleared faster in KO mice than in controls (n = 5, *P < 0.05, **P < 0.005). B: Plasma insulin (μg/L) during ipGTTs is higher (n = 10, *P < 0.05, **P < 0.005). C: Blood glucose after intraperitoneal injection of 0.5 units/kg insulin is similar between wild type and KO (n = 7–8, P = NS). D: Blood glucose (D) and plasma insulin (E) after intravenous injection of glucose (1 g/kg). Both wild-type and KO groups have a broadly similar glucose clearance and insulin level (n = 5–6, P = NS). Blood glucose (F) and plasma insulin (G) after intravenous injection of glucose (1 g/kg) supplemented with the cholinergic analog carbachol (0.53 μmol/L). Compared with controls, KO mice present an improved glucose clearance, associated with a more potent insulin secretion (n = 5–6, *P < 0.05, **P < 0.005).
with anti-Nogo-A antibody was higher than in control antibody–treated animals (control 13.94 ± 5.02 μg/L, 11C7 32.28 ± 8.14 μg/L; n = 3, P < 0.05), yet the total pancreatic insulin content remained unchanged (not shown). No effect was detected on glucose levels or body weight, most probably due to the described severe insulin resistance in peripheral tissues in db/db mice, and also perhaps to the shortness of the treatment period. Antibody-treated db/db mice underwent different metabolic challenges, as follows. During ipGTTs, db/db mice treated with anti-Nogo-A had higher plasma insulin (15-min control 1.25 ± 0.08 μg/L, 11C7 2.17 ± 0.15 μg/L; n = 3–4, P < 0.05) (Fig. 5A). In another experiment, we assessed the glucose-induced insulin secretion by intravenously providing a bolus of glucose (1 g/kg); in this situation, plasma insulin levels were similar most of the time in anti-Nogo-A–treated mice, like in Nogo-A KO mice (Fig. 2E and Fig. 5B). Yet when the mice were given glucose and carbachol (0.53 μmol/L) simultaneously, insulin secretion was fourfold higher on average in anti-Nogo-A–treated db/db animals (control 10.65 ± 2.14, 11C7 46.43 ± 8.00; n = 3, P < 0.05) (Fig. 5C) while the total pancreatic insulin content remained steady (not shown).

We reasoned that the higher insulin secretion observed in db/db mice treated with anti-Nogo-A neutralizing antibody could result from a higher parasympathetic input and/or from higher β-cell cholinergic sensitivity, as in Nogo-A KO mice. We therefore indirectly assessed the parasympathetic tone after 2DG-induced neuroglycopenia by measuring the plasma levels of PP, as above. Interestingly, fasting basal PP levels before 2DG stimulation are comparable between healthy wild-type animals and control db/db mice (not shown), yet after 2DG injection, they become twofold higher in control db/db animals (wild type 101.08 ± 11.45 pg/mL, control db/db 252.78 ± 46.56 pg/mL). In anti-Nogo-A–treated db/db mice, 2DG injection
did not alter the fasting basal plasma PP level (\(n = 5, P = \text{NS}\)) (Fig. 5D). Similarly, like in Nogo-A KO mice, GIP levels were not affected by 2DG administration in random-fed anti-Nogo-A–treated \(db/db\) mice (Fig. 5E) (\(n = 5, P = \text{NS}\)). Altogether, these results suggest that the antibody-mediated neutralization of Nogo-A does not affect the vagal and incretin input on \(\beta\)-cells.

In parallel studies, anti–Nogo-A antibody was given to healthy control mice (C57BL/6J background). This treatment did not affect plasma levels of glucose or insulin (Supplementary Fig. 4), thus addressing a safety issue. The fact that 11C7 treatment has no effect on wild-type mice may be because they have normal parasympathetic input, contrary to Nogo-A KO and 11C7-treated \(db/db\) mice, which have a higher parasympathetic tone (Fig. 4A and Fig. 5D).

These observations were completed with the study of \(\beta\)-cell responsiveness to various secretagogues using isolated \(db/db\) islets cultured in the presence of glucose supplemented with 100 \(\mu\)mol/L carbachol. Like the Nogo-A KO islets (above, Fig. 4B), islets from anti-Nogo-A–treated \(db/db\) mice displayed higher insulin secretion in the presence of glucose supplemented with carbachol (\(n = 7-15, P < 0.05\), yet their sensitivity to glucose supplemented with GLP-1 was not affected (Fig. 5F).

Combined together, these different observations reveal that the administration of Nogo-A–neutralizing antibody enhances insulin secretion in diabetic \(db/db\) mice. Contrary to Nogo-A KO animals, the parasympathetic input on islets is not altered in anti-Nogo-A–treated \(db/db\) animals. However, as in Nogo-A KO mice, \(\beta\)-cell sensitivity to

![FIG. 5. Insulin secretion is improved in diabetic \(db/db\) mice treated with anti–Nogo-A neutralizing antibody (11C7). A–C: Plasma insulin levels (\(\mu\)g/L) after injection of glucose are higher in mice treated with anti–Nogo-A antibody than in controls. A: Intraperitoneal injection of 2 g/kg glucose (\(n = 2–4, *P < 0.05\)). B: Intravenous injection of glucose (1 g/kg) (\(n = 3, *P < 0.05\)). C: Intravenous injection of glucose (1 g/kg) supplemented with carbachol (CC; 0.53 \(\mu\)mol/L); the potentiation effect on insulin secretion is marked in anti-Nogo-A–treated mice (\(n = 3, *P < 0.05\)). D and E: Intravenous injection of 2DG (50 mg/kg), which triggers the parasympathetic stimulation. D: The plasma PP levels (pg/mL) in anti-Nogo-A–treated \(db/db\) mice fasted overnight are similar to those of controls (\(n = 5, P = \text{NS}\)). E: The plasma GIP levels (pg/mL) in random-fed, anti-Nogo-A–treated \(db/db\) mice are similar to those of controls (\(n = 5, P = \text{NS}\)). F: Sensitivity of \(\beta\)-cells from isolated islets of control and 11C7-treated \(db/db\) mice in response to glucose supplemented with CC (10 \(\mu\)mol/L) or GLP-1 (100 nmol/L). Isolated islets from anti-Nogo-A–treated \(db/db\) mice secrete more insulin in response to 4.2 mmol/L + CC and higher glucose concentrations (\(n = 7-15\) batches of 10 islets, four mice, **\(P < 0.005\)).]
glucose supplemented with carbachol is increased in db/db mice after Nogo-A downregulation.

Globally, the observations made with two independent experimental systems suggest that when Nogo-A activity is deleted or significantly impaired, the cholinergic vagal β-cell stimulation triggers a more potent release of insulin.

**β-Cell replenishment after near-total β-cell loss is not improved by downregulation of Nogo-A.** Nogo-A is a negative modulator of regeneration in the CNS (23–25). We therefore explored if it has a similar function in regenerating an injured pancreas. We treated Nogo-A KO mice also bearing the RIP-DTR transgene with diphtheria toxin to induce the near-complete destruction of β-cells, as previously described (36). The trends in glycemia in these mice were followed for up to 3 months, reflecting the lack of any beneficial effect for the loss of Nogo-A (Supplementary Fig. 5A). Accordingly, the total pancreatic insulin content and β-cell mass were unaffected by the downregulation of Nogo-A, thus revealing that Nogo-A inactivation does not improve per se the regeneration capacity of the pancreas after extreme β-cell loss (Supplementary Fig. 5B and C), even though the genetic background of Nogo-A KO mice may somewhat influence the extent of recovery, as reported to occur for spinal cord regeneration (34).

**DISCUSSION**

We report here that 1) Nogo-A is expressed in islet cells and that 2) its genetic inactivation results in increased insulin secretion through vagal stimulation due to a higher parasympathetic input and a higher responsiveness of β-cells to carbachol in the absence of Nogo-A. Similarly, 3) antibody-mediated neutralization of Nogo-A in diabetic db/db mice increases the vagal-induced insulin secretion, by increasing the sensitivity of β-cells to cholinergic stimuli. Finally, 4) contrary to what is observed in cells of the injured CNS, Nogo-A downregulation does not ameliorate the rate of β-cell regeneration in a model of total β-cell ablation.

The high vagal stimulation of insulin secretion was previously reported in obese patients (47,48) and in rodent models of T2D (fa/fa rats, preobese ob/ob mice, high fat–fed mice, and mice subjected to long-term glucose infusion or streptozotocin treatment) (49–54). Yet the precise involvement of the parasympathetic and sympathetic tones, and the sensitivity of β-cells, was not assessed in these studies. Using the high-fat–fed model, Ahren et al. (50) showed higher β-cell sensitivity to carbachol-induced insulin secretion in vitro.

**β-Cell hypersensitivity in Nogo-A KO mice is independent of the glucose-stimulated insulin secretion pathway.** The molecular mode of action of Nogo-A in the pancreas is an intriguing question and deserves further study and elucidation. The changes at the parasympathetic-to-β-cell synapse, possibly via the intracellular messengers of Nogo (Rho and Ca2+), as well as effects on pre- or postsynaptic cytoskeleton, are mechanisms that could be studied in dissociated coculture systems. Perhaps the signal transduction leading to insulin secretion is also altered; we found by quantitative PCR and deep sequencing on isolated islets that the relative β-cell expression of muscarinic and nicotinic receptors, which determine the sensitivity to cholinergic stimulation (55), as well as that of other cell signaling mediators, was identical in extracts from wild-type and Nogo-A KO isolated islets (not shown). Interestingly, the expression of Trpm2, a Ca2+ channel, and serotonin receptor Htr7, well known to potentiate insulin secretion, was upregulated (46,56). We did not explore the activation of phospholipase C and the generation of IP3 and diacylglycerol; however, we observed a similar secretory response to glucose or to glucose supplemented with KCl in isolated wild-type and Nogo-A KO islets (not shown). This suggests that, in the absence of Nogo-A, the improved insulin secretion in response to glucose/carbachol does not rely on the cell machinery involved in glucose/KCl-induced insulin secretion. Such dissociation in the sensitivity to different secretagogues was previously shown in other mutants, like those expressing a dominant-negative form of HNF1α specifically in β-cells (38,50,57).

**Differential involvement of the parasympathetic input.** Insulin secretion is enhanced in diabetic db/db mice when Nogo-A activity is downregulated with antibody treatment; even though this effect is not mediated by a higher parasympathetic input to islets, it is nevertheless enhanced by it. However, the short exposure to anti–Nogo-A antibody (only two injections) was sufficient to increase the sensitivity of β-cells to glucose concentrations >4.2 mmol/L supplemented with carbachol. This observation alone raises new hopes in the quest for finding new ways to promote insulin secretion in diabetic patients.

The phenotype of 1IC7-treated db/db mice partially recapitulates that of Nogo-A KO animals. It must be reiterated, though, that it is unclear whether intravenously injected monoclonal antibodies can cross the blood-brain barrier (58,59). In particular, it was shown that anti–Nogo-A antibody does not cross it (60); therefore, this would prevent any effect on the parasympathetic input from the CNS. However, we cannot completely exclude that a significant action of Nogo-A takes place outside the islets.

Further exploration would be needed to fully address this aspect, such as by generating a β-cell–specific Nogo-A KO mouse. Yet, since Nogo-A downregulation also results in improved insulin secretion in vitro, from isolated islets, we conclude that Nogo-A acts locally (i.e., at the islet level). Together, these observations are compatible with Nogo-A action through combined mechanisms (intra- and extras- islet).

In conclusion, we have reported herein that Nogo-A is involved in glucose homeostasis. These results suggest that Nogo-A is a potential new target for antidiabetic drugs, by promoting insulin secretion in response to cholinergic stimuli, such as after food intake, without hypoglycemic events. We must recall, in this regard, that a neutralizing anti-human Nogo-A antibody is currently being tested in phase I clinical trials on acutely injured paraplegic patients in centers of the European Network of Spinal Cord Injury (M.E.S., in collaboration with Novartis). The inhibition of Nogo-A, for instance, with a weekly injection of antibody may thus represent an avenue for new antidiabetic treatments, by acting on the stimulatory input and thereby on insulin secretion from β-cells.

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